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1 **The utility of anti-Müllerian hormone in women with chronic kidney disease, on**  
2 **haemodialysis and after kidney transplantation**

3 **Short title: Anti-Müllerian hormone in women with renal failure**

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21 **Key Message**

22 Anti-Müllerian hormone (AMH) is lower in young women with renal failure compared with  
23 age-matched healthy controls but not in those on haemodialysis. AMH decreases with age in  
24 women with renal failure in a similar manner to the general population. AMH may have a  
25 role as a marker of ovarian health in non-dialysis renal patients pursuing pregnancy.

26 **Abstract**

27 Women with renal disease have menstrual and gonadal dysfunction manifesting as hormonal  
28 imbalance. Anti-Müllerian hormone (AMH) has been described as a potential measure of the  
29 ovarian reserve. We examined circulating AMH levels in young women with renal failure  
30 divided into 3 groups, determined associations with clinical characteristics, and compared  
31 AMH with age-matched healthy individuals. AMH concentrations were measured in 77  
32 women (mean age  $32.9 \pm 5.4$  years); 26 had chronic kidney disease (CKD) stages 3-5, 26 were  
33 on haemodialysis (HD), and 25 had a kidney transplant. Random AMH levels are highest in  
34 women on HD [HD 2.9 (1.1-5.2), CKD 1.6 (0.7-2.2), transplant 1.5 (1.0-4.2)ng/mL]. On  
35 multiple linear regression, AMH was 53% higher (95% CI 0.20-0.98,  $p=0.002$ ) in women on  
36 HD and decreased by 20% per 5-year increase in age ( $p<0.001$ ). AMH was 43% lower in  
37 women with renal failure compared with 600 age-matched controls [1.7 (0.9-3.8) vs 3.0 (1.9-  
38 5.0)ng/mL,  $p<0.001$ ]; however, we found no difference in AMH between those on HD and  
39 healthy individuals [2.9 (1.1-5.2) vs 3.0 (1.9-5.0)ng/mL]. AMH may be a useful biomarker in  
40 female renal patients with non-dialysis dependent renal disease pursuing pregnancy. In  
41 contrast, AMH levels are higher in HD but unlikely to reflect ovarian reserve.

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43 **Keywords:** Anti-Müllerian hormone; chronic kidney disease; haemodialysis; kidney  
44 transplant; fertility.

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51 **Introduction**

52 Anti-Müllerian hormone (AMH) is a glycoprotein with a fundamental role in male sex  
53 differentiation. In women, AMH plays a critical role in folliculogenesis, with circulating  
54 levels directly reflecting the number of developing preantral follicles and indirectly the  
55 number of primordial follicles in the ovaries (Iliodromiti et al., 2015). As such, AMH is now  
56 recognised as the best available biomarker of both the functional and true ovarian reserve  
57 (Dewailly et al., 2014). Accurate quantitative assessment of the ovarian reserve by AMH  
58 (Anderson et al., 2015) has enabled prediction of reproductive lifespan, tailoring of fertility  
59 preservation and optimisation of assisted conception outcomes (Dewailly et al., 2014; Nyboe  
60 Andersen et al., 2016). The recent development of a fully automated Elecsys® AMH  
61 immunoassay (Gassner and Jung 2014) with enhanced sensitivity, specificity and  
62 reproducibility, has widened its clinical utility and enabled assessment of women with limited  
63 ovarian function.

64 Women with advanced chronic kidney disease (CKD) often have disturbances in the  
65 menstrual cycle and amenorrhea is common by the time the patient reaches end stage renal  
66 disease (ESRD) (Zingraff et al., 1982). The menstrual cycle typically remains irregular even  
67 after the initiation of maintenance dialysis. Consistent with this, pregnancy is extremely  
68 uncommon as one progresses from CKD stage 3 to dialysis (Zingraff et al., 1982).  
69 Conversely, fertility is frequently restored within a few months after successful kidney  
70 transplantation (Levidiotis et al., 2009). To date we are only aware of a single small study  
71 (n=60) assessing AMH in patients with renal failure(Sikora-Grabka et al., 2016). This study  
72 utilised a manual AMH ELISA, which was limited by complement interference,  
73 irreproducible results and limited sensitivity (Iliodromiti et al., 2015).

74 The aim of this study was to measure serum AMH concentrations in predialysis, dialysis and  
75 kidney transplant women of childbearing age; explore potential factors affecting AMH and  
76 compare AMH levels with age-matched healthy controls.

77

## 78 **Materials and methods**

### 79 **Design and Participants**

80 This was a single-centre cohort study of all women aged 18-40 years attending renal services  
81 between August 1, 2015 and March 31, 2016 in our catchment area (serving a population of  
82 approximately 1.5 million). Potential participants were identified from the electronic patient  
83 record used in our centre and by screening clinic lists. A letter was sent to all eligible patients  
84 to make them aware of the research, which included an opt in or opt out reply slip where they  
85 could suggest a way for the research staff to contact them to discuss the study further. If they  
86 opted in, they were contacted by a member of the research team to discuss the details of the  
87 study and organise a study visit. The protocol of the study was approved by the Research  
88 Ethics Committee (REC reference: 15/NS/0040) on 20<sup>th</sup> May 2015. The study was conducted  
89 in accordance with the Declaration of Helsinki, and written informed consent was obtained  
90 from all participants.

91 We measured serum AMH levels in three distinct groups of patients; women with CKD  
92 stages 3-5, women on haemodialysis (HD), and kidney transplant recipients. We excluded  
93 individuals with active or previous cancer (breast, ovarian, lymphoma, pelvic radiotherapy),  
94 ovarian surgery, current use of alkylating agent-based protocols, severe active illness and  
95 patients with inability to provide informed consent.

96 AMH levels from a multicentre study using the same assay in 600 age-matched healthy  
97 women with regular menstrual cycles, not on contraception were used as the reference group  
98 (Anckaert et al., 2016).

**100 Baseline data**

101 Demographics, aetiology of renal failure, duration of renal replacement therapy (RRT), actual  
102 day of the menstrual cycle during the examination, gynaecological history [including  
103 menstrual characteristics, number of pregnancies or miscarriages, history of polycystic ovary  
104 syndrome (PCOS), family history of premature menopause], and medications potentially (but  
105 not definitively) related to AMH concentrations (contraceptives (Bentzen et al., 2012; Deb et  
106 al., 2012), prednisolone (Ubaldi et al., 2002) and cyclophosphamide (Clowse et al., 2011))  
107 were recorded.

108 In addition to AMH, we measured a number of other hormones regulating the ovarian  
109 function, including follicle-stimulating hormone (FSH), luteinizing hormone (LH), prolactin,  
110 oestradiol and progesterone. Also, serum creatinine, C-reactive protein (CRP), and thyroid  
111 hormones blood concentrations were analysed based on literature showing potential  
112 associations with AMH (Yarde et al., 2014; Polyzos et al., 2015; Weghofer et al., 2016).

113 Menstrual cycles were defined as regular when menstrual flow occurred every 21 to 35 days,  
114 irregular when menstrual flow occurred less than 21 days or more than 35 days apart, and  
115 amenorrhoea was defined as the abnormal absence of menstruation for 90 days or more in  
116 accordance with the NICE guideline (NICE 2014). The Chronic Kidney Disease  
117 Epidemiology Collaboration (CKD-EPI) equation was used for calculation of estimated  
118 glomerular filtration rate (eGFR) (Levey et al., 2009).

119

**120 Study procedures**

121 The blood samples were collected on a random day during the menstrual cycle and then  
122 anonymised and centrifuged. In the women on HD, blood samples were obtained before a HD  
123 session.

124 For measurement of AMH, 3mL serum aliquots for each patient were stored at -80°C. AMH  
125 was measured on first thaw of stored samples using an automated method on a clinically  
126 validated platform (e411, Roche Diagnostics, Burgess Hill, UK) (Gassner and Jung 2014).  
127 The assay was calibrated and quality controlled using the manufacturer's reagents. Detection  
128 limit was 0.01ng/mL and the coefficient of variation between runs for two levels of control  
129 ran at <8%. All AMH samples (including controls) were measured in the same laboratory by  
130 the same laboratory-developed test methods in a single run, and all values can therefore be  
131 compared uniformly.

132 All other biochemical parameters were measured using standard assays, in a National Health  
133 Service clinical biochemistry department.

134

### 135 **Statistical analysis**

136 We examined differences in demographic, clinical factors, and biochemical parameters  
137 stratified by renal failure group. Continuous variables were expressed with means and  
138 standard deviations (SD) or medians and interquartile ranges (for non-parametric data), and  
139 analysed using parametric and non-parametric tests as appropriate. Categorical variables were  
140 reported as frequencies and percentages and proportions compared by chi-squared or Fisher's  
141 exact tests.

142 For all comparisons, values of AMH were log transformed to normalise their distribution and  
143 were analysed as continuous variables.

144 One-way ANOVA was performed in the transformed data to examine the percentage change  
145 in AMH between renal failure groups. Means (and SD) obtained from the logged values were  
146 exponentiated (back transformed) and are therefore the geometric means, which were derived  
147 after subtraction of the constant.

148 Correlation analyses were performed to assess the relationship between log transformed  
149 AMH and each predictor variable. Variables tested were age, weight, alcohol and smoking  
150 history, eGFR, renal failure group, cause of established renal failure (ERF), biochemical  
151 parameters (reproductive hormones, thyroid hormones and CRP), day of menstrual cycle,  
152 presence or not of menstrual periods, and medications potentially related to follicular growth  
153 (contraceptives, steroids, cyclophosphamide). Factors with p value <0.10 in correlation  
154 analyses were tested in linear regression models with log transformed AMH as the outcome  
155 variable. Coefficients obtained from linear regression models of the logged values were  
156 exponentiated (back transformed) and are therefore ratios of geometric means per  
157 unit/category change of the exposure and should be interpreted as proportional change per  
158 exposure with a null value of 1.

159 Mean log transformed AMH levels of young women with renal failure were compared with  
160 age-matched healthy references using a two-sample z-test. Z-test was calculated for all  
161 patients and in subgroups stratified by age ( $\leq 25$ , 26-30, 31-35, and  $\geq 36$  years) and by renal  
162 failure group (CKD stages 3-5, HD, and transplant).

163 The study was designed to enroll 75 participants (25 in each renal failure group), which  
164 would be sufficient to detect a significant difference between the healthy controls and women  
165 with renal failure assuming the difference in AMH between a healthy and renal failure  
166 woman is 0.4ng/mL and SD in each group is 0.7ng/mL, providing power of 80% and  
167 probability of type 1 error of 5%(Morel et al., 2013).

168 For all analyses, a p value <0.05 was considered significant. The IBM SPSS Statistics  
169 Package (version 21.0; SPSS, Inc., Armonk, NY) was used for all analyses.

170

## 171 **Results**

### 172 **Study population**



173 We measured AMH levels in 77 women of childbearing age who were attending the renal  
174 services and fulfilled the study entry criteria. Twenty-six had CKD stages 3-5, 26 were on  
175 HD, and 25 had a kidney transplant (Table 1).

176 Mean age of the enrolled participants was 32.9 (SD 5.4) years, 38 (49.3%) had regular  
177 menstrual periods, and approximately a third were on hormonal contraception. More than half  
178 had previous successful pregnancies and 18 (23.4%) had at least one miscarriage in the past.  
179 Six women (7.8%) had a history of PCOS. No differences of clinical importance were  
180 identified between the three groups (Table 1).

181

### 182 **Biochemical parameters**

183 Median AMH was higher in HD patients (2.9, IQR 1.1-5.2ng/mL) compared with CKD  
184 stages 3-5 patients (1.6, IQR 0.7-2.2ng/mL) and transplant patients (1.5, IQR 1.0-4.2ng/mL)  
185 (Table 2). By comparing the logged AMH values by ANOVA, there was significant  
186 difference in AMH levels between renal failure groups [ $F(2, 74)=3.686, p=0.03$ ]. Tukey post  
187 hoc analysis revealed that AMH was significantly higher in HD patients ( $p=0.02$ ) compared  
188 with CKD stages 3-5 patients. There was no difference between HD and transplant patients or  
189 between CKD and transplant patients. With regard to the rest of the biochemical parameters,  
190 there were no differences of note between the three groups, other than thyroid stimulating  
191 hormone (TSH) being marginally higher in the CKD group (although still within the 'normal'  
192 range) and the transplant patients having better renal function than those with CKD as  
193 expected.

194 From all variables tested, age and renal failure group were associated with AMH on  
195 regression analysis (Table 3). When multiple linear regression analysis was performed, there  
196 was an average 4% (95%CI 0.02 to 0.06,  $p<0.001$ ) decrease in AMH level per year increase  
197 in age when accounting for the renal failure group (Table 3). When compared with women

198  $\leq 25$  years, women older than 36 years had 39% (95%CI 0.08 to 0.62,  $p=0.04$ ) lower AMH  
199 levels (Table 3). Patients on HD had higher AMH levels by 53% (95%CI 0.20 to 0.98,  
200  $p=0.002$ ) compared with CKD patients, following adjustment for age. No differences were  
201 found between the other renal failure groups. The multiple regression model fit was R-  
202 squared=0.24 (Table 3). There was no difference in AMH levels between women with  
203 menstrual periods vs amenorrhoeic (1.7, IQR 0.8-3.9ng/mL vs 1.5, IQR 1.0-3.0ng/mL,  
204 respectively), previously treated with cyclophosphamide vs non-treated (1.5, IQR 0.4-  
205 1.5ng/mL vs 1.7, IQR 0.9-3.9ng/mL, respectively), and women on hormonal contraception vs  
206 not (1.6, IQR 1.1-3.8ng/mL vs 1.7, IQR 0.8-3.4ng/mL, respectively).

207 Women with renal failure had 43% lower AMH levels than healthy women (1.7, IQR 0.9-  
208 3.8ng/mL vs 3.0, IQR 1.9-5.0ng/mL,  $p<0.001$ ) (Figure 1). In subgroup analysis, AMH was  
209 lower in all age groups in women with renal failure, apart from those aged 26-30 years  
210 ( $\leq 25$ yr 1.7 vs 4.0ng/mL,  $p<0.04$ ; 26-30yr 3.7 vs 3.2ng/mL, not significant; 31-35yr 2.1 vs  
211 2.6ng/mL,  $p=0.004$ ;  $\geq 36$ yr 1.0 vs 1.7ng/mL, not significant). When stratified by renal failure  
212 group, in comparison with healthy age-matched women controls, AMH was lower in women  
213 with CKD stages 3-5 (1.6 vs 3.0ng/mL,  $p<0.001$ ) and women with a kidney transplant (1.5 vs  
214 3.0ng/mL,  $p<0.001$ ) but not in women on HD (2.9 vs 3.0) (Figure 2).

215

## 216 **Discussion**

217 The data suggest that women of childbearing age with CKD stages 3-5 or a kidney transplant  
218 have lower AMH compared with age-matched women without kidney disease and this may  
219 contribute to the low fertility rate in this population. Notably, women on haemodialysis had  
220 similar AMH levels compared with age-matched controls and this may reflect an intrinsic  
221 dysregulation of the granulosa cells leading to higher AMH production or alternatively AMH  
222 accumulation in ERF requiring dialysis. In women with renal disease, increasing age was

223 associated with a reduction in AMH concentrations similar to that observed in the general  
224 population. Although pregnancy is extremely uncommon in this patient population, AMH  
225 may have a role as a marker of ovarian health in non-dialysis female renal patients pursuing  
226 pregnancy. In haemodialysis patients AMH levels seem to be inappropriately high therefore,  
227 patients in this group should be excluded from conclusions drawn about the relationship  
228 between AMH and ovarian reserve.

229 To date, AMH has been developed with a wide array of clinical applications (Nelson 2013;  
230 Dewailly et al., 2014). These include prediction of the ovarian response to stimulation with  
231 exogenous gonadotrophins for in-vitro fertilisation, the duration of the reproductive lifespan  
232 and diagnosis of premature ovarian insufficiency, disorders of sex development and PCOS  
233 (Seifer et al., 2002; Stubbs et al., 2005; Josso et al., 2012; Iliodromiti et al., 2013; Rey et al.,  
234 2013; Tehrani et al., 2013; La Marca and Sunkara 2014). AMH has also been used for the  
235 assessment of gonadotoxicity of cancer therapy, monitoring of granulosa cell tumors to detect  
236 residual or recurrent disease, and assessing the loss of the ovarian tissue secondary to ovarian  
237 surgery (La Marca and Volpe 2007; Geerts et al., 2009; Anderson and Wallace 2013).  
238 Women with renal failure and childbearing potential are a diverse group of patients with  
239 complex pathologies where AMH may provide valuable insight, especially to those  
240 considering future pregnancy.

241 About 3% of women of childbearing age are affected by renal disease (Williams and Davison  
242 2008). However, the incidence of pregnancy in women with CKD stages 3-5 is difficult to  
243 determine as it is not routine practice to measure kidney function in pregnant women in the  
244 United Kingdom, unless there is some other indication. Furthermore, kidney function is  
245 difficult to interpret during pregnancy as GFR increases. The frequency of conception among  
246 women of childbearing age undergoing RRT ranges from 0.3 to 1.5% per year (Holley et al.,  
247 1997). For transplant patients the unadjusted pregnancy rate is 33-45 per 1000 women (Gill et

248 al., 2009; Stoumpos et al., 2016) compared with more than 100 per 1000 women in the  
249 general population.

250 Circulating AMH concentrations reflect the functional ovarian reserve and indirectly the  
251 number of residual primordial follicles within the ovary. As a consequence of this intimate  
252 relationship with the ovarian reserve, a decline in AMH may indicate both physiological and  
253 premature aging of the gonads (Kalaiselvi et al., 2012; Younis 2012). Applicable to the  
254 population with renal failure, in women with moderate to severe CKD (stages 3-5), the risk of  
255 complications to mother and fetus with pregnancy is high enough that some advocate against  
256 pregnancy or to postpone until they receive a kidney transplant. Similarly, female kidney  
257 transplant recipients are traditionally counselled to wait one to two years after transplantation  
258 before conceiving (McKay and Josephson 2008). This postponement frequently leads to  
259 women with renal disease attempting to have children during a period where female fertility  
260 is already in decline due to ageing. Furthermore, menopause in women with renal failure  
261 occurs 4.5 years earlier than in healthy women (Weisinger and Bellorin-Font 2004) leaving  
262 them with fewer potential childbearing years. Nonetheless, not all cases are clear-cut and  
263 there is a number of women with either advanced renal disease or a kidney transplant in  
264 whom assessment of AMH may be useful to guide pregnancy planning. For example, a young  
265 woman with normal AMH values would potentially have more time to deal with any  
266 underlying medical issues. In contrast, women with a borderline or low AMH, indicative of a  
267 diminished ovarian reserve, may be counselled that fertility preservation may be  
268 advantageous.

269 Women on HD were found to have higher AMH concentrations compared with the other two  
270 renal failure groups and this was an unexpected finding. Impaired glomerular filtration could  
271 be a potential mechanism although AMH has a molecular weight of 140kD, which is too  
272 large to cross the basement membrane of the glomerular capillaries. On the other hand, AMH

273 is of similar size to other molecules perceived to be uraemic ‘toxins’ (Duranton et al., 2012)  
274 (i.e. fibrinogen,  $\alpha$ 2-macroglobulin) and although their concentrations are not directly  
275 associated with glomerular function they are increased in dialysis, so accumulation of AMH  
276 in dialysis is plausible. Uraemia is known to interfere with the metabolism and regulation of a  
277 number of endogenous hormones; however, the direct impact of uraemia in ovarian follicles  
278 and AMH levels is not clear. We hypothesise that high AMH levels during dialysis are  
279 probably related to the follicular arrest during the selection process of the dominant follicle,  
280 through a negative interaction between AMH and FSH (Grossman et al., 2008). If so, AMH  
281 significantly decreases FSH-induced oestradiol production, which leads to disruption of  
282 normal antral follicular development and maturation, similarly to what is happening in  
283 women with PCOS (Agarwal et al., 1996). In accordance with this, oestradiol levels were  
284 lower in women on HD compared with the other two renal failure groups, although the  
285 difference was not significant. In a study of 186 young healthy Danish women (Hagen et al.,  
286 2012) that were followed until they conceived or for six menstrual cycles, the fecundability  
287 ratio (FR) (i.e., the monthly probability of conceiving) in women with low AMH levels was  
288 similar to women with medium AMH levels (FR 0.81; 95% CI 0.44–1.40) but women with  
289 high AMH levels had reduced FR (FR 0.62; 95% CI 0.39–0.99). This supports our findings,  
290 where high AMH levels may represent women with conditions of anovulation. In a recently  
291 published study (Sikora-Grabka et al., 2016), women on HD had similar AMH concentrations  
292 with healthy women and this is similar to our findings. However, when women on HD were  
293 stratified into those with regular menstrual cycles vs not, the regularly menstruating women  
294 on HD had significantly lower serum AMH concentration compared with the healthy  
295 controls. In our study there was no difference in AMH concentrations between women on HD  
296 with regular, irregular periods or amenorrhoea; however, the numbers were very small (n=11,  
297 n=8 and n=7, respectively). In the same study, AMH concentrations were found to gradually

298 decrease from pre- to 6 months post-transplantation and this may reflect the transition from  
299 abnormally high AMH concentrations during dialysis to lower ‘normal’ concentrations post-  
300 transplantation.

301 Our study has a number of strengths including the use of a cohort of young, well-phenotyped  
302 women with renal failure, distinct clinical categorisation of disease progression and that we  
303 could compare them with a large cohort of age-matched healthy controls. We performed  
304 extensive biochemical characterisation, including use of a robust automated AMH assay. We  
305 do however, acknowledge some limitations: information on menstrual cycle characteristics  
306 were not easy to ascertain in all cases, especially in women with scanty menstrual periods;  
307 however, AMH has been shown to be stable throughout the menstrual cycle and in women  
308 with irregular cycles random sampling is inevitable. We had limited power to assess the  
309 effects of cyclophosphamide or the combined oral contraceptive pill, but overall AMH was  
310 reduced in keeping with previous larger studies. In women with renal failure other markers of  
311 ovarian reserve, such as antral follicle count (AFC) which is an ultrasound biomarker of  
312 follicle number may be a better predictor of oocyte yield than AMH. We were unable to  
313 perform transvaginal ultrasound examination and AFC measurement to provide additional  
314 assessment of the ovarian reserve; however, AFC is not routinely available and is known to  
315 exhibit substantial intra- and interobserver variability. Lastly, we accept that no causal  
316 inferences on the effect of renal failure on AMH levels can be made, with prospective  
317 longitudinal studies best placed to address this.

318 We demonstrate that in women of reproductive age with CKD stages 3-5 or a renal  
319 transplant, AMH exhibits a similar age-related decline but age-specific values are lower than  
320 equivalent healthy controls. For women on HD circulating AMH concentrations are higher,  
321 suggesting a disruption of folliculogenesis or impaired clearance. AMH evaluation may be a  
322 useful biochemical test in estimating the reproductive lifespan in women with renal disease

323 not treated with dialysis, pursuing preconception counselling. However, it is important to  
324 note that AMH is only a marker of ovarian reserve and should be used as an adjuvant tool.  
325 Conception can be achieved with low AMH levels therefore, one should be careful about the  
326 interpretation of AMH as a prognostic marker for conception success. Further research is  
327 warranted to investigate the direct impact of uraemia on ovarian follicles and AMH  
328 production.

329

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331 Dr Martin Hund is an employee of Roche Diagnostics. All other authors report no conflicts of  
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333

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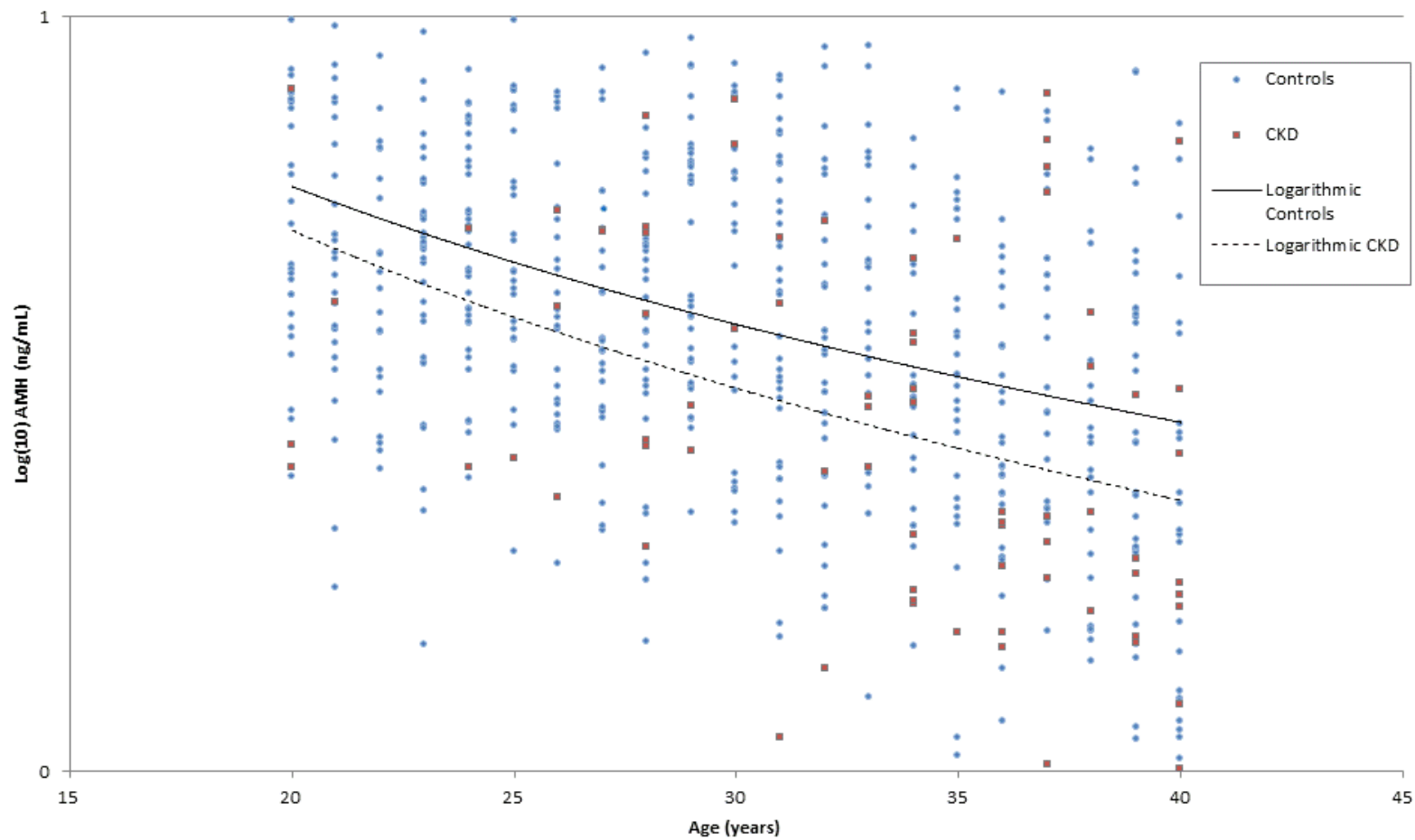
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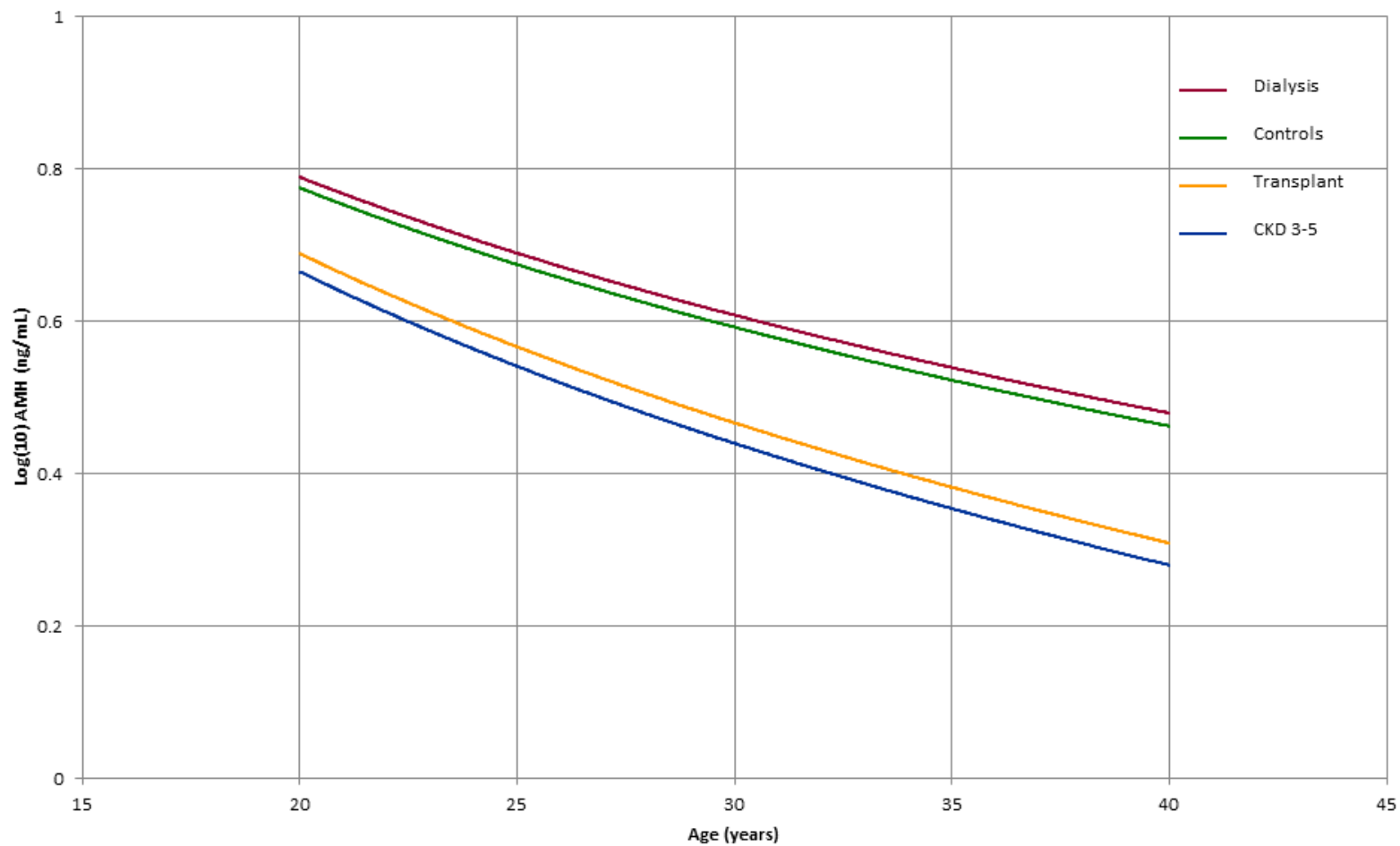
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487 **Figure 1.** Log AMH levels in young women with CKD (n=77) compared with AMH levels in a cohort of 600 age-matched healthy women  
488 (p<0.001, two-sample z-test). Data are log-transformed.  
489 AMH, anti-Müllerian hormone; CKD, chronic kidney disease.



490

491 **Figure 2.** Age-related distribution of serum log AMH levels in women of reproductive age with CKD stages 3-5 (n=26), on HD (n=26), kidney  
492 transplant recipients (n=25) and healthy controls (n=600). Data are log-transformed.  
493 AMH, anti-Müllerian hormone; CKD, chronic kidney disease; HD, haemodialysis.



494

|  | All patients (n=77) | CKD (n=26)  | HD (n=26)      | Transplant (n=25) | <i>p-value</i> <sup>a</sup> |
|--|---------------------|-------------|----------------|-------------------|-----------------------------|
| Age (years), mean (SD)                                     | 32.9 (5.4)          | 33.4 (5.4)  | 34.0 (4.9)     | 31.2 (5.8)        | NS                          |
| Weight (kg), mean (SD)                                     | 68.1 (17.8)         | 71.2 (18.8) | 66.8 (15.3)    | 66.1 (19.4)       | NS                          |
| Ethnic origin  |                     |             |                |                   |                             |
| Caucasian, n (%)   | 66 (85.7)           | 20 (76.9)   | 26 (100.0)     | 20 (80.0)         | 0.04                        |
| Asian, n (%)   | 11 (14.3)           | 6 (23.1)    | 0 (0.0)        | 5 (20.0)          |                             |
| Smoking <sup>b</sup> , n (%)                               | 25 (32.5)           | 7 (26.9)    | 12 (46.2)      | 6 (24.0)          | NS                          |
| Alcohol <sup>c</sup> , n (%)                               | 40 (51.9)           | 17 (65.4)   | 11 (42.3)      | 12 (48.0)         | NS                          |
| Cause of ERF   |                     |             |                |                   |                             |
| Glomerulonephritis, n (%)                                  | 31 (40.3)           | 10 (38.5)   | 9 (34.6)       | 12 (48.0)         | NS                          |
| Congenital renal dysplasia/reflux, n (%)                   | 14 (18.2)           | 3 (11.5)    | 5 (19.2)       | 6 (24.0)          |                             |
| Diabetes, n (%)  | 12 (15.6)           | 6 (23.1)    | 5 (19.2)       | 1 (4.0)           |                             |
| Other <sup>d</sup> , n (%)                                 | 20 (26.0)           | 7 (26.9)    | 7 (26.9)       | 6 (24.0)          |                             |
| Years on RRT, median (IQR)                                 | 4.3 (1.6, 11.3)     |             | 2.8 (0.6, 7.5) | 8.7 (3.1, 12.0)   | 0.03                        |
| Menstrual periods  |                     |             |                |                   |                             |
| Amenorrhoea, n (%)   | 15 (19.5)           | 5 (19.2)    | 7 (26.9)       | 3 (12.0)          | NS                          |
| Regular, n (%)   | 38 (49.4)           | 12 (46.2)   | 11 (42.3)      | 15 (60.0)         |                             |
| Irregular, n (%)   | 24 (31.2)           | 9 (34.6)    | 8 (30.8)       | 7 (28.0)          |                             |
| Previous pregnancies, n (%)                                | 46 (59.7)           | 15 (57.7)   | 17 (65.4)      | 14 (56.0)         | NS                          |
| Previous miscarriages, n (%)                               | 18 (23.4)           | 8 (30.8)    | 5 (19.2)       | 5 (20.0)          | NS                          |
| Polycystic ovary syndrome, n (%)                           | 6 (7.8)             | 2 (7.7)     | 1 (3.8)        | 3 (12.0)          | NS                          |
| Family history of premature menopause <sup>e</sup> , n (%) | 3 (5.5)             | 3 (16.7)    | 0 (0.0)        | 0 (0.0)           | 0.04                        |
| Medications related to follicular growth                   |                     |             |                |                   |                             |
| Hormonal contraception, n (%)                              | 29 (37.7)           | 12 (46.2)   | 8 (30.8)       | 9 (36.0)          | NS                          |
| Steroids, n (%)  | 37 (48.1)           | 5 (19.2)    | 8 (30.8)       | 24 (96.0)         | <0.001                      |
| Previous cyclophosphamide, n (%)                           | 5 (6.5)             | 2 (7.7)     | 1 (3.8)        | 2 (8.0)           | NS                          |

<sup>a</sup> ANOVA or chi-squared test or Fisher's exact test or Mann-Whitney U test where appropriate

<sup>b</sup> Current or previous vs never

<sup>c</sup> Occasionally (<14units/week) vs never

<sup>d</sup> Polycystic kidney disease (n=6), unknown aetiology (n=6), haemolytic uremic syndrome (n=2), post-acute kidney injury (n=2), malignant hypertension (n=1), tubulointerstitial nephritis (n=1), amyloidosis (n=1), nephronophthisis (n=1)

<sup>e</sup> Excludes 22 patients with missing values

CKD, chronic kidney disease; HD, haemodialysis; SD, standard deviation; NS, not statistically significant; ERF, established renal failure; RRT, renal replacement therapy; IQR, interquartile range.

| <b>Table 2. Biochemical parameters stratified by renal failure group</b>   |                     |                   |                   |                   |                             |
|--|---------------------|-------------------|-------------------|-------------------|-----------------------------|
|  | All patients (n=77) | CKD (n=26)        | HD (n=26)         | Transplant (n=25) | <i>p-value</i> <sup>a</sup> |
| eGFR <sup>b</sup> , mL/min/1.73m <sup>2</sup> ; mean (SD)  | 50.8 (27.6)         | 34.4 (15.3)       |                   | 67.2 (27.7)       | <0.001                      |
| <45 <sup>b</sup> , n (%)   | 26 (51.0)           | 19 (73.1)         |                   | 7 (28.0)          | 0.005                       |
| ≥45, n (%)   | 25 (49.0)           | 7 (26.9)          |                   | 18 (72.0)         |                             |
| Day of menstrual cycle <sup>c</sup> , median (IQR)   | 12 (6-22)           | 16 (5-24)         | 12 (4-22)         | 12 (9-17)         | 0.77                        |
| Anti-Müllerian hormone, ng/mL; median (IQR)  | 1.7 (0.9, 3.8)      | 1.6 (0.7, 2.2)    | 2.9 (1.1, 5.2)    | 1.5 (1.0, 4.2)    | 0.03 <sup>d</sup>           |
| Anti-Müllerian hormone, ng/mL; geometric mean (SD) <sup>e</sup>  | 1.9 (0.7)           | 1.4 (0.5)         | 2.6 (0.8)         | 1.8 (0.8)         |                             |
| Follicle-stimulating hormone, IU/L; median (IQR)   | 4.1 (2.5, 6.0)      | 5.1 (3.7, 6.0)    | 3.8 (2.6, 5.6)    | 3.0 (2.5, 6.3)    | 0.37                        |
| Luteinising hormone, IU/L; median (IQR)  | 6.5 (3.8, 10.0)     | 6.8 (3.8, 8.3)    | 5.0 (3.6, 11.5)   | 5.6 (3.8, 12.4)   | 1.00                        |
| Prolactin, mIU/L; median (IQR)   | 360 (247, 563)      | 306 (243, 469)    | 444 (345, 652)    | 315 (206, 544)    | 0.08                        |
| Oestradiol, pmol/L; median (IQR)   | 242 (102, 395)      | 226 (113, 326)    | 130 (82, 317)     | 320 (132, 585)    | 0.12                        |
| Progesterone, nmol/L; median (IQR)   | 1.3 (1.0, 4.7)      | 1.1 (1.0, 4.8)    | 1.6 (1.2, 4.2)    | 1.0 (1.0, 3.2)    | 0.45                        |
| Thyroid stimulating hormone, mIU/L; median (IQR)   | 1.5 (0.9, 2.1)      | 1.8 (1.4, 2.4)    | 1.3 (0.9, 1.7)    | 1.1 (0.7, 2.0)    | 0.03                        |
| Free thyroxine, pmol/L; median (IQR)   | 13.2 (12.4, 14.9)   | 12.9 (12.3, 14.6) | 13.2 (12.4, 14.1) | 14.0 (12.7, 15.5) | 0.38                        |
| C-reactive protein, mg/L; median (IQR)   | 2.5 (1.0, 6.3)      | 2.0 (1.0, 5.0)    | 3.5 (1.0, 9.8)    | 2.0 (1.0, 5.0)    | 0.37                        |
| <sup>a</sup> T-test or chi-squared test or Kruskal-Wallis test or ANOVA where appropriate<br><sup>b</sup> Excludes patients on dialysis<br><sup>c</sup> Excludes 15 women with amenorrhoea and 8 women with menstrual cycles >35 days<br><sup>d</sup> P-value was obtained by comparing logged values<br><sup>e</sup> Geometric means were derived from logged AMH values after back transformation<br>CKD, chronic kidney disease; HD, haemodialysis; eGFR, estimated glomerular filtration rate; SD, standard deviation; IQR, interquartile range. |                     |                   |                   |                   |                             |

| <b>Covariate</b>           | <b>Simple linear regression</b>                   |                |                 | <b>Multiple linear regression</b>                 |                |                 |
|----------------------------|---|----------------|-----------------|---|----------------|-----------------|
|                            | <b>Ratio of geometric means (Bootstrap 95%CI)</b> | <b>p-value</b> | <b>R-square</b> | <b>Ratio of geometric means (Bootstrap 95%CI)</b> | <b>p-value</b> | <b>R-square</b> |
| Age (each additional year) | -0.04 (-0.06 to -0.02)                            | 0.001          | 0.15            | -0.04 (-0.06 to -0.02)                            | <0.001         | 0.24            |
| Renal failure group        |   |                | 0.09            |   |                |                 |
| CKD (reference)            | 1.0   | -              |                 | 1.0   | -              |                 |
| HD                         | 0.49 (0.16 to 1.00)                               | 0.009          |                 | 0.53 (0.20 to 0.98)                               | 0.002          |                 |
| Transplant                 | 0.16 (-0.13 to 0.53)                              | 0.31           |                 | 0.06 (-0.18 to 0.36)                              | 0.70           |                 |
| Age group                  |   |                | 0.16            |   |                |                 |
| ≤25 (reference)            | 1.0   | -              |                 |   |                |                 |
| 26-30                      | 0.04 (-0.34 to 0.60)                              | 0.88           |                 |   |                |                 |
| 31-35                      | -0.20 (-0.49 to 0.22)                             | 0.36           |                 |   |                |                 |
| ≥36                        | -0.39 (-0.62 to -0.08)                            | 0.04           |                 |   |                |                 |

AMH, anti-Müllerian hormone; CI, confidence interval; CKD, chronic kidney disease; HD, haemodialysis.