

Research Paper



2025; 16(7): 2075-2086. doi: 10.7150/jca.105590

A novel prognostic model to predict prognosis of patients with osteosarcoma based on clinical characteristics and blood biomarkers

Shulin Chen^{1,2}, Liru Tian^{1,3}, Chuan Li¹, Dongmei Zhong¹, Tingting Wang¹, Yuyu Chen⁴, Taifeng Zhou⁴, Xiaoming Yang⁴, Zhiheng Liao⁴, Caixia Xu^{1⊠}

- 1. Research Center for Translational Medicine, the First Affiliated Hospital, Sun Yat-sen University, 58 Zhongshan Road 2, Guangzhou, Guangdong 510080, P.R. China.
- 2. Department of Clinical Laboratory, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou 510060, P. R. China.
- Department of Pathology, State Key Laboratory of Oncology in South China, Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou 510060, P. R. China.
- 4. Guangdong Provincial Key Laboratory of Orthopedics and Traumatology, Department of Spine Surgery, the First Affiliated Hospital of Sun Yat-sen University, Guangzhou 510080, China.

Shulin Chen and Liru Tian have contributed equally to this work.

🖂 Corresponding author: Caixia Xu. xucx3@mail.sysu.edu.cn.

© The author(s). This is an open access article distributed under the terms of the Creative Commons Attribution License (https://creativecommons.org/licenses/by/4.0/). See https://ivyspring.com/terms for full terms and conditions.

Received: 2024.10.21; Accepted: 2025.02.16; Published: 2025.03.10

Abstract

Purpose: Osteosarcoma (OSC) is a high-morbidity bone cancer with an unsatisfactory prognosis. Timely and accurate assessment the overall survival (OS) and progression-free survival (PFS) in patients with OSC are required to guide and select the best treatment. This study aimed to develop a simple, convenient and low-cost prognostic model based on clinical characteristics and blood biomarkers for predicting OS and PFS in OSC patients.

Methods: Overall, 158 patients with OSC included from the Sun Yat-sen University Cancer Center in this retrospective study. LASSO-Cox algorithm was used to shrink predictive factor size and established a prognostic risk model for predicting OS and PFS in OSC patients. The predictive ability of the survival model was compared to the Tumor Node Metastasis (TNM) stage and clinical treatment by concordance index (C-index), time-dependent receiver operating characteristic (td-ROC) curve, decision curve analysis (DCA), net reclassification improvement index (NRI), and integrated discrimination improvement index (IDI).

Results: Based on results from the LASSO-Cox method, gender, family history of cancer, monocyte (M), red blood cell (RBC), lactic dehydrogenase (LDH), and cystatin C (Cys-C) were identified to construct a novel predictive model for the OSC patients. The C-index of the prognostic model to predict OS and PFS were 0.713 (95% CI = 0.630 - 0.795) and 0.636 (95% CI = 0.577 - 0.696), respectively, which were higher than the OS and PFS of TNM stage and clinical treatment. Td-ROC curve and DCA of the predictive model also demonstrated good predictive accuracy and discriminatory power of OS and PFS compared to TNM stage and treatment. Moreover, the prognostic model performed well across all time frames (1-, 3-, and 5-year) with regards to the IDI and NRI in comparison to the TNM stage, and clinical treatment.

Conclusion: The simple, convenient and low-cost prognostic model we developed demonstrated favorable performance for predicting OS and PFS in OSC patients, which may serve as a useful tool for physicians to provide personalized survival prediction for OSC patients.

Keywords: Osteosarcoma; Blood biomarkers; LASSO-Cox regression; Prognostic

Introduction

Osteosarcoma (OSC) is the most common bone cancer, as well as one of the most common primary malignancies among children and adults[1, 2]. The incidence of OSC is 2–3/million/year in the general population, though epidemiologic statistics indicate that the incidence and mortality of OSC have been increasing at approximately 1.4% per year[3]. OSC mostly develops in the long bone near the epiphyseal growth plate of the extremities. The most common sites include the distal femur, proximal tibia, and proximal humerus. OSC is characterized by a high tendency to metastasize and local recurrence[4]. The incidence ratio of male to female is 1.4[5].

Despite multidisciplinary therapies, including surgical excision, radiotherapy, and chemotherapy[6], many OSC patients still experience tumor recurrence and metastasis, which results in poor prognosis and low survival rates among this group of patients[2, 7]. The presence or absence of metastasis, local recurrence, chemotherapy regimen, chemotherapy response, patient characteristics, tumor staging, tumor characteristics, and neoadjuvant tumor cell destruction percentage has an effect on prognosis[1, 7]. Specifically, metastasis has the greatest impact on prognosis. The overall survival rate of patients with metastatic disease is only about 20 - 30%, compared to 70 - 80% for non-metastatic patients[8].

Clinical staging is a common method of assessing risk of OSC[9]. However, clinical staging systems for OSC, which include the American Joint Committee on Cancer (AJCC) staging and Enneking staging are only able to provide a rough assessment of the clinical risk for OSC based on pathological grade, tumor size, and metastasis. However, survival differs among patients with the same tumor stage[10]. These results indicate that the traditional staging system is inadequate for predicting the survival of cancer patients without considering other prognostic factors (such as clinical characteristics or blood biomarkers). Therefore, it is necessary to explore more reliable prognostic indicators to remedy shortcomings of the staging system, as well as to improve the prediction of clinical outcomes for patients with OSC.

Blood-based liquid biopsy has emerged as a useful tool for diagnosis and the prediction of outcome in patients with cancer. A variety of blood biomarkers have been studied with regards to the diagnosis and follow-up of OSC progression and recurrence, including alkaline phosphatase (ALP) and lactate dehydrogenase (LDH). Among the different blood biomarkers tested, ALP has been shown to have the most diagnostic value for OSC, and has been shown to be positively correlated with tumor volume, which has an additional useful prognostic significance[11]. Thus far, the clinical characteristics in combination with blood-based biomarkers were used to predict a prognosis of OSC had little been reported.

Thus, the retrospective study aimed to construct a prognostic model based on clinical characteristics and blood biomarkers for predicting the overall survival (OS) and progression-free survival (PFS) in patients with OSC, as well as to assess its incremental predictive ability in traditional TNM stage and clinical treatment of individual OS and PFS.

Materials and methods

Patient selection and data collection

Patients hospitalized and treated at the Sun Yat-sen University Cancer Center (SYSUCC) between January 2010 and December 2019 were consecutively enrolled into this present retrospective study. The inclusion criteria were as follows: (1) histologically confirmed OSC; (2) patients did not receive any anti-cancer treatment; (3) cancer-specific survival: alive or dead due to cancer. The exclusion criteria: (1) the existing malignancy other than OSC, alone or in combination with OSC; (2) Incomplete clinical information, laboratory data, and follow-up data. This study was granted approval by the Clinical Research Ethics Committee of the Sun Yat-sen University Cancer Center(B2022-481; Date of Approval: August 5, 2022). Since the study was retrospective in nature, the Institutional Review Board waived written informed consent. In addition, this study was carried out according to the principles of the Declaration of Helsinki.

Baseline clinical characteristics were collected from the patients' medical records, which included age, gender, smoking status, family history, tumor site, tumor size, tumor border, clinical treatment, and the eighth edition of the American Joint Committee on Cancer (AJCC) tumor-node-metastasis (TNM) staging[12]. The pretreatment blood-routine biomarkers were collected within one week before administering anti-cancer treatment. The biomarkers that were collected included white blood cell (WBC), neutrophil (N), lymphocyte (L), monocyte (M), platelet (PLT), neutrophil / lymphocyte ratio (NLR), lymphocyte / monocyte ratio (LMR), platelet/ lymphocyte ratio (PLR), derived neutrophil-tolymphocyte ratio (dNLR), prognostic nutritional index (PNI)[13], red blood cell (RBC), hemoglobin (HGB), serum phosphorus (IP³⁺), serum calcium $(Ca^{2+}),$ serum magnesium (Mg²⁺), alanine aminotransferase (ALT), aspartate aminotransferase (AST), AST / ALT ratio (SLR), alkaline phosphatase (ALP), lactic dehydrogenase (LDH), glutamyl transpeptidase (GGT), total protein (TP), albumin (ALB), globulin (GLOB), C-reactive protein (CRP), ALB / CRP ratio (ACR), total bile acid (TBA), urea, creatinine (CRE), cystatin C (Cys-C), uric acid (UA), total cholesterol (CHO), triglycerides (TG), high density lipoprotein cholesterol (HDL-C), low density lipoprotein cholesterol (LDL-C), LDL-C / HDL-C ratio (LHR), apolipoprotein AI (APOA), apolipoprotein B (APOB), APOA / APOB ratio (ABR), glucose (GLU).

Patients follow up

Data relevant to patient demographics and laboratory test data were abstracted from the electronic medical record. Follow-up was done by telephone or outpatient service, and the deadline for follow-up was April 2022. Overall survival (OS) was measured from the date of diagnosis until death due to cancer or the end point of the study. Progression-free survival (PFS) was calculated from date of the objective disease progression or death or the date of the last follow-up.

Statistical analysis

Patients' characteristics were shown as frequencies (percentages) for categorical variables and mean ± standard deviation (SD) for continuous variables. We used the t-test or Wilcoxon test to compare mean. LASSO-Cox regression algorithm was adopted to select the most useful prognostic factors related to OS and constructed a novel prognostic model. Subsequently, Harrell's C-index (C-index), time-dependent ROC (td-ROC) curves[14], decision analysis (DCA)[15], net reclassification curve improvement index (NRI), and integrated discrimination improvement index (IDI)[16] were used to compare the prognostic performances of the novel prognostic model with TNM stage and clinical treatment. DCA was utilized to evaluate clinical validity of the prognostic model and quantifying the net benefits at different threshold probabilities [17]. The NRI assessed the ability of a new model to reclassify subjects into binary event or no-event categories compared to an older model. The IDI index quantified the improvement in average sensitivity without reducing the average specificity of a new model compared to the older model[18]. The correlation between the novel prognostic model, TNM stage, and clinical treatment was assessed using by Pearson's correlation coefficient. In addition, we constructed a nomogram that integrates the prognostic model risk score, TNM stage, and clinical treatment that may assist in individual survival prediction of OSC patients. Internal validation and calibration of the nomogram were performed via

bootstrap resampling procedure. Finally, according to the risk score, OSC patients were classified into low-risk groups and high-risk groups, Kaplan-Meier method was used to compare the two groups in terms of PFS and OS. Statistical analyses were performed with R programing language (version 3.6.1) and Graph Pad Prism (version 5.0, San Diego, CA, USA), the results were considered statistically significant if the P value of less than 0.05.

Results

Demographic characteristics of the enrolled patients

Overall, 158 patients with osteosarcoma (OSC) were included in this retrospective study. 92 (58.2%) of these patients were male, and 66 (41.8%) were female. The median age was 16 years (95% confidence interval [CI], 15-18). According to 8th edition of the AJCC TNM stage criteria, the number of patients in stages I&II and III&IV was 121 (76.6%) and 37 (23.4%), respectively. The median OS was 32.5 months (95% CI, 30.4-36.2), The median PFS was 23.3 months (95% CI, 17.9-28.7). The baseline characteristics of the total OSC patients were shown in Table 1.

The correlation between clinical characteristics and blood biomarkers

The correlation between clinical characteristics and blood biomarkers were shown in supplement table 1. The numbers in the table shown the Pearson's correlation coefficient between different variables. We found that different clinical characteristics were significantly associated with different blood biomarkers. Age was the most correlated with blood biomarkers, which significantly correlated with 20 (PLT, IP³⁺, ALT, SLR, ALP, GGT, Urea, CRE, UA, Cys-C, LDH, GLU, TG, CHO, HDL-C, LDL-C, LHR, APOA, APOB, and ABR) out of 39 blood biomarkers.

Establishment of a prognosis model for OS and PFS

Firstly, by using the LASSO-Cox regression analysis (Figure 1A), the optimal lambda value (lambda = 0.054) was obtained via minimum criteria (Figure 1B), and its corresponding six predictors (gender, family history of cancer, monocyte, RBC, Cys-C, and LDH) were correlated with OS. Finally, a simple prognostic model consisting of the six predictors were screened out in the LASSO-Cox regression analysis and were then generated risk score based on the regression coefficients. The prognostic model risk score was calculated using the following formula: risk score = $-(0.0247 \times gender) (0.6294 \times family history of cancer) - (0.5652 \times monocyte)$ - (0.1119×RBC) + (0.0019 ×LDH) + (0.1293×Cys-C). In this formula, the following dichotomous variables: gender (Male=1, Female=0), family history of cancer (yes=1, no=0), the other continuous variables value represents their respective serum levels. This formula was applied to calculate each patient's risk score.

Table	1.	Demographics	and	clinical	characteristics	of	OSC
patients							

Variable	No. (%) or mean ±standard deviation (SD)
Characteristics	
Age (years)	22.1 ± 15.4
Gender	
Male	92 (58.2%)
Female	66 (41.8%)
Smoke	
Yes	7 (4.4%)
No	151 (95.6%)
Family history of cancer	
Yes	9 (5.7%)
No	149 (94.3%)
Tumor site	
Skull	19 (12.0%)
Trunk	8 (5.1%)
Extremities	131 (82.9)
Treatment	
NAC plus Sur plus Che	105 (66.5%)
Sur plus Che	8 (5.1%)
Sur	17 (10.8%)
Che	25 (15.8%)
Other	3 (1.8%)
TNM stage ^a	
I&II	121 (76.6%)
III&IV	37 (23.4%)
Laboratory data	
WBC (10 ⁹ /L)	8.18 ± 3.19
Neutrophil (10 ⁹ /L)	5.53 ± 2.96
Lymphocyte (10 ⁹ /L)	1.99 ± 0.66
Monocyte (10 ⁹ /L)	0.49 ± 0.25
PLT (10 ⁹ /L)	310.45 ± 83.19
NLR	3.11 ± 2.48
LMR	5.46 ± 8.13
PLR	171.34 ± 72.56
dNLR	2.29 ± 2.05
PNI	54.69 ± 6.46
$\frac{RBC}{RBC} (10^{12}/L)$	4 79 + 0.65
$\frac{HGB}{(\sigma/L)}$	131 20 + 19 61
$\frac{1100 (g/2)}{100}$	1 39 + 0 23
$\frac{1}{Ca^{2+} (\text{mmol}/\text{L})}$	2 40 + 0.68
$\frac{\mathrm{Cu}^{2+}(\mathrm{mmol}/\mathrm{L})}{\mathrm{Mg}^{2+}(\mathrm{mmol}/\mathrm{L})}$	0.90 ± 0.08
	16.98 + 12.57
$\frac{\text{AET}(U/L)}{\text{AST}(U/L)}$	20.93 + 8.34
SLR	1 70 + 1 14
	470 73 + 1038 01
	277 24 + 241 36
	23 20 + 16 33
$\frac{O(1/L)}{TP(\alpha/L)}$	74 31 + 6 15
$\frac{11 (g/L)}{AIB(\alpha/L)}$	<u>14.71 + 5.42</u>
1100 (8/ 0)	TT./ 1 1 J.TJ

Variable	No. (%) or mean ±standard deviation (SD)
CRP (mg/L)	10.64 ± 20.24
ACR	52.34 ± 112.58
TBA (umol/L)	4.98 ± 5.67
Urea (mmol/L)	4.40 ± 1.51
CRE (umol/L)	52.83 ± 17.91
Cys-C (mg/L)	0.80 ± 0.16
UA (umol/L)	338.15 ± 93.32
CHO (mmol/L)	4.06 ± 1.01
TG (mmol/L)	1.19 ± 0.65
HDL-C (mmol/L)	1.19 ± 0.30
LDL-C (mmol/L)	2.46 ± 0.89
LHR	2.19 ± 0.95
APOA (g/L)	1.21 ± 0.23
APOB (g/L)	0.80 ± 0.23
ABR	1.63 ± 0.53
GLU (mmol/L)	5.17 ± 1.00

a: TNM stage was classified according to the AJCC 8th TNM staging system; Abbreviations: TNM: Tumor Node Metastasis; NAC: neoadjuvant therapy; Sur: surgery; Rad: radiotherapy; Che: chemotherapy; PLT: platelet; NLR: neutrophil / lymphocyte ratio; LMR: lymphocyte / monocyte ratio; PLR: platelet / lymphocyte ratio; dNLR: derived neutrophil-to-lymphocyte ratio; PNI: prognostic nutritional index; RBC: red blood cell; HGB: hemoglobin; IP³⁺: serum phosphorus; Ca²⁺: serum calcium; Mg²⁺: serum magnesium; ALT: alanine aminotransferase; AST: aspartate aminotransferase; SLR: AST / ALT ratio; ALP: alkaline phosphatase; LDH: lactic dehydrogenase; GGT: glutamyl transpeptidase; TP: total protein; ALB: albumin; CRP: C-reactive protein, ACR: ALB / CRP ratio; TBA: total bile acid; CRE: creatinine; Cys-C: cystatin C; UA: uric acid; CHO: total cholesterol; TG: triglycerides; HDL-C: high density lipoprotein cholesterol; LDL-C: low density lipoprotein cholesterol; LHR: LDL-C / HDL-C ratio; APOA: apolipoprotein A1; APOB: apolipoprotein B; ABR: APOA / APOB ratio; GLU: glucose.

The comparison between the prognostic model with TNM stage and clinical treatment

The C-index, td-ROC curve, DCA, NRI, and IDI were used to compare the prognostic performances of the novel prognostic model with TNM stage and clinical treatment. Firstly, the C-index was calculated and compared to the C-index of the three predictive signatures (Table 2). For OS, the C-index of the prognostic model was 0.713 (95% CI = 0.630-0.795), which was significantly higher than that in TNM stage [0.590 (95% CI = 0.518 - 0.663), P = 0.011] and clinical treatment [0.604 (95% CI = 0.521-0.687), P = 0.020]. With regards to PFS, the C-index of the novel prognostic model was 0.636 (95% CI = 0.577 - 0.696), which was shown to be significantly higher compared to that of the TNM stage [0.552 (95% CI = 0.503-0.600), P = 0.022] and clinical treatment [0.517 (95% CI = 0.461–0.574), *P* = 0.001]. Secondly, the td-ROC curves were plotted, and the area under ROC curves (AUCs) of the three predictive signatures were calculated. For OS, the AUCs of 1-, 3-, and 5-year were 0.783, 0.730, and 0.743, respectively. And for PFS, the AUCs of 1-, 3-, and 5-year were 0.701, 0.674, and 0.658, respectively. The AUCs of the novel prognostic model were higher compared to that of TNM stage and clinical treatment, with regards to both OS and PFS at 1-, 3-, and 5-year. (Figure 2). Thirdly, the DCA demonstrated that within the most reasonable



Figure 1. Potential predictors selection using LASSO-Cox regression analysis. (A) The changing trajectory of each variable in LASSO-Cox regression analysis; (B) Selection of the optimal lambda value with the minimum partial likelihood deviance.

threshold probability range of OS and PFS, the novel prognostic model had a higher overall net benefit than the TNM stage and clinical treatment (Figure 3). Finally, both NRI and IDI calculations were obtained at 1-, 3-, and 5-year, and were utilized to compare alternative prognostic indices of our prognostic model with TNM stage and clinical treatment. Positive value represents better accuracy while negative value represents worse accuracy. The results were presented in Table 3. For OS, NRI analysis indicated that the prognostic model had higher predictive power compared to that of the TNM stage and clinical treatment both at 1-, 3-, and 5-year OS survival. IDI analysis also indicated that that the discrimination ability of the novel prognostic model was higher than that of the TNM stage and clinical treatment. In addition, similar results also indicated that the novel prognostic model had improved performance in predicting PFS for OSC patients than others.

Table 2. The C-index of OS and PFS for prognostic model, TNMstage, and treatment.

Survival prediction	C-index	95 CI%	Р			
For OS						
Prognostic model	0.713	0.630 - 0.795				
TNM stage	0.590	0.518 - 0.663				
Treatment	0.604	0.521 - 0.687				
Prognostic model vs TNM stage			0.011			
Prognostic model vs Treatment						
For PFS						
Prognostic model	0.636	0.577 - 0.696				
TNM stage	0.552	0.503 - 0.600				
Treatment	0.517	0.461 - 0.574				
Prognostic model vs TNM stage			0.022			
Prognostic model vs Treatment						

C-index = concordance index; P values are calculated based on normal approximation using function rcorrp.cens in Hmisc package.

Establishment of predictive nomogram

The nomogram incorporated the prognostic model risk score, TNM stage, and clinical treatment to quantitative analysis the 1-, 3-, and 5- years OS (Figure 4A) and PFS (Figure 4B) survival probability for each OSC patient. The points of the factors indicate their corresponding contribution to survival probability. And the calibration curves indicated the nomogram-predicted 1-, 3-, 5-year OS (Figure 4C) and PFS (Figure 4D) were matched well with actual 1-, 3-, 5-year OS and PFS.

Performance of the prognostic model in stratifying risk

According to the risk scores, the patients were classified into low-risk groups and high-risk groups, differences in survival between the two groups were tested using the Kaplan-Meier method and compared by the log rank tests. Patients in the high-risk group (risk score \geq -0.27) tended to have a worse OS (Figure 5A; P < 0.001) and PFS (Figure 5B; P < 0.001) than those in the low-risk group (risk score < -0.27). Additionally, we wanted to test whether the prognostic model would be able to make up for the current deficiencies of the AJCC TNM stage and clinical treatment. Next, patients were factitiously stratified into the early stage (stage I/II), the late stage (stage III/IV), and received the same clinical treatment (neoadjuvant therapy plus surgical resection plus chemotherapy). Kaplan-Meier curve indicated that high-risk patients in the early stage had significantly lower OS (Figure 5C; P = 0.010) and PFS (Figure 5F; P = 0.003) compared to low-risk patients.

In the late stage, the OS (Figure 5D; P < 0.001) and PFS (Figure 5G; P = 0.007) in the low-risk and high-risk groups also displayed significant difference. In the patients received the same clinical treatment, the result suggested a poorer prognosis in the high-risk group both in OS (Figure 5E; P < 0.001) and PFS (Figure 5H; P < 0.001).

Differences between the high-risk and low-risk group in the 6 selected predictors

Figure 6 showed the composition of the low-risk

and high-risk patients by gender and family history of cancer, and compared serum values of monocyte, RBC, LDH, and Cys-C between the two groups. There was significant statistical difference in gender, monocyte, RBC, and LDH between low-risk and high-risk patients. Monocyte and RBC levels in low-risk were higher than the high-risk group.

Table 3. A comparison of discriminatory ability of prognostic model with TNM stage and treatment using NRI and IDI for OS and PFS.

	1-Year			3-Year				5-Year				
	NRI	Р	IDI%	Р	NRI	Р	IDI	Р	NRI	Р	IDI	Р
For OS												
Prognostic model vs TNM stage	0.378	0.149	0.200	< 0.01	0.202	0.060	0.100	0.030	0.129	0.308	0.077	0.308
Prognostic model vs Treatment	0.344	0.239	0.194	0.040	0.151	0.239	0.081	0.209	0.418	0.01	0.131	0.020
For PFS												
Prognostic model vs TNM stage	0.193	0.100	0.058	0.060	0.161	0.209	0.054	0.109	0.063	0.557	0.042	0.318
Prognostic model vs Treatment	0.345	0.060	0.076	< 0.01	0.253	0.090	0.075	0.010	0.280	0.269	0.054	0.209



Figure 2. Time dependent ROC curves analysis of the novel prognostic model, TNM stage, and clinical treatment in OS and PFS. (A-C) Time-dependent AUC curves of the prognostic model, TNM stage, and clinical treatment at 1-year OS, 3-year OS, and 5-year OS; (D-F) Time-dependent AUC curves of the prognostic model, TNM stage, and clinical treatment at 1-year PFS, 3-year PFS.



Figure 3. Decision curve analysis of the novel prognostic model, TNM stage, and clinical treatment in OS and PFS. (A) Decision curve analysis of OS; (B) Decision curve analysis of PFS.



Figure 4. The nomogram plots for estimating OS (A) and PFS (B) at 1, 3, and 5 years. Calibration curves of the nomogram for OS (C) and PFS (D). NSC: neoadjuvant therapy + surgery + chemotherapy; SC: surgery + chemotherapy; Che: chemotherapy.



Figure 5. Kaplan-Meier (K-M) curves with log-rank p-values of the prognostic model. (A, B) K-M curves for OS and PFS of low-risk and high-risk patients basing on the prognostic model; (C, D) K-M curves of OS prognostic value for the prognostic model in the full groups with tumor stage; (E) K-M curves of OS prognostic value for the prognostic model in the full groups with clinical treatment; (F, G) K-M curves of PFS prognostic value for the prognostic model in the prognostic model in the full groups with clinical treatment; (F, G) K-M curves of PFS prognostic value for the prognostic model in the full groups with clinical treatment. NAC: neoadjuvant therapy; Sur: surgery; Che: chemotherapy.



Figure 6. Differences between the high-risk and low-risk group in the gender, family history of cancer, monocyte, RBC, LDH, and Cys-C. (A) gender; (B) family history of cancer; (C) monocyte; (D) RBC; (E) LDH; (F) Cys-C.





Correlation analysis between the prognostic model, TNM stage, and clinical treatment

To investigate the relationship between the prognostic model, TNM stage, and clinical treatment, a Pearson correlation coefficient (PCC) analysis was performed (Figure 7A), where the red represented negative correlation and the blue represented positive correlation, and the circle size represented the size of correlation coefficient. The results indicated that the prognostic model was significantly and positively correlated with TNM stage (PCC = 0.26, P = 0.001), as well as clinical treatment (PCC = 0.19, P = 0.018). In

addition, prognostic model, TNM stage, clinical treatment, and status were shown in the Sankey plot and there was a positive regulatory relationship between prognostic model, TNM stage, clinical treatment, and status (Figure 7B).

Discussion

In this research, we focused on the relationship between the clinical characteristics and blood biomarkers and prognostic value for patients with OSC. The LASSO-Cox regression method was applied to build a model to predict OS and PFS in OSC patients. Our prognostic model was used to predict OS and PFS in OSC patients with a high accuracy and stability, which outperformed the traditional TNM stage and clinical treatment.

Based on results from the LASSO-Cox regression, six prognostic factors (gender, family history of cancer, monocyte, RBC, LDH, and Cys-C) were screened out and then incorporated into the novel predictive model for OSC patients. It had been reported that gender, monocyte, and LDH were related to prognosis of OSC, while levels of RBC and Cys-C were known to be related to the prognosis of other malignant tumors. Monocyte played significant roles in the establishment of the immune microenvironment of OSC. The higher monocyte levels could inhibit the metastasis of OSC, and had longer overall survival times[19, 20]. The serum level of LDH was known to reflect systemic cancer burden, growth, survival and invasive potential[21]. Previous studies had shown that serum LDH level was associated with the prognosis of OSC patients[22, 23]. Family history of cancer was associated with developing cancer, furthermore, it was recognized as an important prognostic factor in several cancers[24-27]. RBC level was one of the RBC parameters, and a reduced preoperative RBC count may reflect poor liver function, which could affect survival of patient survival[28, 29]. Lu et al. reported that in patients with primary liver cancer, patients whose preoperative RBC counts were lower than normal demonstrated a lower OS rate compared to patients whose preoperative RBC counts did not decrease[30]. Cys-C was an endogenous marker of glomerular filtration rate, numerous studies had shown that abnormal serum Cys-C levels can be used as a prognostic and diagnostic indicator for several malignancies[31-34]. All of the data indicated that these prognostic factors were closely related to tumor development, and suggested that our analysis had reliable prognostic value.

In order to evaluate whether our model was able to remedy the deficiencies of the TNM stage and clinical treatment in the prognostic assessment of OSC patients, the OSC patients were divided into the low-risk and high-risk groups based on their prognostic model risk scores. Kaplan-Meier survival curves showed that the high-risk groups had lower OS and PFS in OSC patients with stage I/II, stage III/IV, and clinical treatment. Thus, the results reminded us that even for patients at the same stage, and received the same clinical treatment, high-risk patients will likely require more intense treatment. Moreover, the results also indicated that our model could remedy deficiencies of the TNM stage, and enhance the predictive power of the TNM stage. Improved prediction of individual prognosis could

aid clinicians in many ways, including consulting patients, choosing personalized treatment, and arrange patient follow-up.

This study had several advantages compared to previous studies[35-37]: Firstly, in the past, most prognostic factors were single indicators, but our study had included more potential prognostic factors than the previous studies. And this prognostic model was established based on clinical characteristics and clinical laboratory blood tests available in most clinical settings. So, it is simple, convenient and low-cost for clinical application, especially the application of grass-roots hospitals. Second, we utilized the new algorithm LASSO-Cox regression analysis to develop a prognostic model as a statistical method to filter variables and establish a prognostic model. This allowed us to adjust the model's overfitting to avoid extreme predictions, thereby significantly improving the prediction accuracy. This method has been applied across many studies[38-40]. Thirdly, in this study, we utilized multiple methods to compare the predictive accuracy and discriminative ability of the novel prognostic model with TNM stage and clinical treatment. In addition, these results all that our model outperformed demonstrated compared to others. Lastly, the endpoint of this study was OS and PFS, so this model could achieve better clinical application.

The research still presented some limitations: 1. Selection bias was unavoidable due to the retrospective design of this research. So, its calculated predictive value was for clinicians' reference only. 2. The sample size was relatively small, single-centre data collection, lack of external validation which limit generalizability of the prognostic model. 3. The model was non-specific prediction tool for OSC patients, and may lack certain specificity. Some specific biomarkers may be incorporated into the prognostic model to improve the specificity, such as immunohistochemical markers[41], radiomics[42, 43], and the recently newly applied non-coding RNAs[44, 45]. 4. The subtype of OSC was reported as a prognostic marker, different subtypes of OSC had different survival outcomes[46]. As this study failed to obtain OSC subtypes for each patient, in was not included in our study as a potential prognostic marker. 5. We collected data only for the initial diagnosis and did not dynamically monitor the entire course of the patient. Thus, we could not know the significance of biomarkers for prognosis of the patient after each treatment. 6. We did not compare our model with previously developed and validated prognostic models. This study was a retrospective study, we cannot obtain the data contained in these models from our electronic medical records. Therefore, we did not compare our prognostic model

with previously developed prognostic models. The AJCC TNM staging system of osteosarcoma is the most widely used method for survival prediction. So, we replaced the previously developed models with AJCC TNM staging system and compare it with our model. Despite these shortcomings, the prognostic model was effective and could help predict prognosis of OSC patients, thus providing clinicians with a more practical and convenient tool for individualized treatment decision making and survival assessment at the initial diagnosis.

Conclusions

In conclusion, we successfully constructed a 6 clinical characteristics and blood biomarkers-based prognostic model for OSC patients. This model outperformed TNM stage and clinical treatment in predicting OS and PFS in OSC patients. Low-cost and satisfactory stability of this prognostic model may act as a useful tool for physicians to provide consultation, personalized survival prediction for OSC patients. But, the properties of our prognostic model required verification before wide variety of clinical applications.

Supplementary Material

Supplementary table. https://www.jcancer.org/v16p2075s1.pdf

Acknowledgements

We thank the staff at the Director of Clinical Laboratories, Sun Yat-sen University Cancer Center for providing support on research conditions in this study.

Funding

This work was supported by the National Natural Science Foundation of China (No. 82472388). Guangdong Province Basic and Applied Basic Research Fund Project (No. 2023A1515010307).

Author contributions

Shulin Chen and Liru Tian had full access to all of the data in the study and take responsibility for the content of the manuscript, including the data and analysis. Chuan Li, Dongmei Zhong, Tingting Wang, Yuyu Chen and Taifeng Zhou contributed to study conception and design. Xiaoming Yang and Zhiheng Liao contributed to acquisition, analysis, or interpretation of data. Caixia Xu is accountable for all aspects of work. All authors reviewed the manuscript.

Ethics approval

This work was approved by the ethics committee

of the Clinical Research Ethics Committee of the Sun Yat-sen University Cancer Center (B2022-481; Date of Approval: August 5, 2022). Since the study was retrospective in nature, the Institutional Review Board waived written informed consent. In addition, this study was carried out according to the principles of the Declaration of Helsinki.

Data availability statement

For reasons of patient privacy, the data sets analyzed in the current study are not publicly available, but it can be obtained from the corresponding author (xucx3@mail.sysu.edu.cn) under reasonable requirements.

Competing Interests

The authors have declared that no competing interest exists.

References

- Ottaviani G, Jaffe N. The epidemiology of osteosarcoma. Cancer treatment and research. 2009; 152: 3-13.
- Friebele JC, Peck J, Pan X, Abdel-Rasoul M, Mayerson JL. Osteosarcoma: A Meta-Analysis and Review of the Literature. American journal of orthopedics (Belle Mead, NJ). 2015; 44: 547-53.
- Pei Y, Yao Q, Li Y, Zhang X, Xie B. microRNA-211 regulates cell proliferation, apoptosis and migration/invasion in human osteosarcoma via targeting EZRIN. Cell Mol Biol Lett. 2019; 24: 48.
- Xing D, Qasem SA, Owusu K, Zhang K, Siegal GP, Wei S. Changing prognostic factors in osteosarcoma: analysis of 381 cases from two institutions. Human Pathology. 2014; 45: 1688-96.
- Stiller CA, Bielack SS, Jundt G, Steliarova-Foucher E. Bone tumours in European children and adolescents, 1978-1997. Report from the Automated Childhood Cancer Information System project. European Journal of Cancer. 2006; 42: 2124-35.
- Ando K, Heymann M-F, Stresing V, Mori K, Heymann D. Current Therapeutic Strategies and Novel Approaches in Osteosarcoma. Cancers. 2013; 5: 591-616.
- Anderson ME. Update on Survival in Osteosarcoma. Orthop Clin North Am. 2016; 47: 283-92.
- Duchman KR, Gao Y, Miller BJ. Prognostic factors for survival in patients with high-grade osteosarcoma using the Surveillance, Epidemiology, and End Results (SEER) Program database. Cancer epidemiology. 2015; 39: 593-9.
- Heck RK, Stacy GS, Flaherty MJ, Montag AG, Simon MA. A comparison study of staging systems for bone sarcomas. Clinical Orthopaedics & Related Research. 2003; 415: 64-71.
- Zhang J, Yang J, Wang HQ, Pan Z, Yan X, Hu C, et al. Development and validation of a nomogram for osteosarcoma-specific survival: A population-based study. Medicine. 2019; 98: e15988.
- 11. Lindsey BA, Markel JE, Kleinerman ES. Osteosarcoma Overview. Rheumatology and therapy. 2017; 4: 25-43.
- Doescher J, Veit JA, Hoffmann TK. The 8th edition of the AJCC Cancer Staging Manual. HNO. 2017; 65: 956-61.
- He X, Li JP, Liu XH, Zhang JP, Zeng QY, Chen H, et al. Prognostic value of C-reactive protein/albumin ratio in predicting overall survival of Chinese cervical cancer patients overall survival: comparison among various inflammation based factors. Journal of Cancer. 2018; 9: 1877-84.
- Kamarudin AN, Cox T, Kolamunnage-Dona R. Time-dependent ROC curve analysis in medical research: current methods and applications. BMC medical research methodology. 2017; 17: 53.
- Talluri R, Shete S. Using the weighted area under the net benefit curve for decision curve analysis. BMC medical informatics and decision making. 2016; 16: 94.
- Burch PM, Glaab WE, Holder DJ, Phillips JA, Sauer JM, Walker EG. Net Reclassification Index and Integrated Discrimination Index Are Not Appropriate for Testing Whether a Biomarker Improves Predictive Performance. Toxicological sciences: an official journal of the Society of Toxicology. 2017; 156: 11-3.
- Vickers AJ, Van Calster B, Steyerberg EW. Net benefit approaches to the evaluation of prediction models, molecular markers, and diagnostic tests. Bmj. 2016; 352: i6.
- Farinholt P, Park M, Guo Y, Bruera E, Hui D. A Comparison of the Accuracy of Clinician Prediction of Survival Versus the Palliative Prognostic Index. Journal of pain and symptom management. 2018; 55: 792-7.

- 20. Chen T, Zhao L. Patrolling monocytes inhibit osteosarcoma metastasis to the lung. Aging. 2020; 12: 23004-16.
- Liu R, Cao J, Gao X, Zhang J, Wang L, Wang B, et al. Overall survival of cancer patients with serum lactate dehydrogenase greater than 1000 IU/L. Tumour biology: the journal of the International Society for Oncodevelopmental Biology and Medicine. 2016; 37: 14083-8.
- Fu Y, Lan T, Cai H, Lu A, Yu W. Meta-analysis of serum lactate dehydrogenase and prognosis for osteosarcoma. Medicine. 2018; 97: e0741.
- 23. Marais LC, Bertie J, Rodseth R, Sartorius B, Ferreira N. Pre-treatment serum lactate dehydrogenase and alkaline phosphatase as predictors of metastases in extremity osteosarcoma. Journal of bone oncology. 2015; 4: 80-4.
- Yuequan J, Shifeng C, Bing Z. Prognostic factors and family history for survival of esophageal squamous cell carcinoma patients after surgery. The Annals of thoracic surgery. 2010; 90: 908-13.
- Xiao R, Wu J, Ward BB, Liu H, Li B, Wang C, et al. Family history of cancer is associated with poorer prognosis in oral squamous cell carcinoma. Oral diseases. 2023; 29: 2066-75.
- Chattopadhyay S, Zheng G, Hemminki A, Försti A, Sundquist K, Sundquist J, et al. Influence of family history on risk of second primary cancers and survival in patients with squamous cell skin cancer. The British journal of dermatology. 2020; 183: 488-94.
- Han MA, Oh MG, Choi IJ, Park SR, Ryu KW, Nam BH, et al. Association of family history with cancer recurrence and survival in patients with gastric cancer. Journal of clinical oncology: official journal of the American Society of Clinical Oncology. 2012; 30: 701-8.
- Naeshiro N, Aikata H, Kakizawa H, Hyogo H, Kan H, Fujino H, et al. Long-term outcome of patients with gastric varices treated by balloon-occluded retrograde transvenous obliteration. Journal of gastroenterology and hepatology. 2014; 29: 1035-42.
- Imai K, Beppu T, Yamao T, Okabe H, Hayashi H, Nitta H, et al. Clinicopathological and prognostic significance of preoperative serum zinc status in patients with hepatocellular carcinoma after initial hepatectomy. Annals of surgical oncology. 2014; 21: 3817-26.
- Xie X, Yao M, Chen X, Lu W, Lv Q, Wang K, et al. Reduced red blood cell count predicts poor survival after surgery in patients with primary liver cancer. Medicine. 2015; 94: e577.
- Mulaomerović A, Halilbasić A, Cickusić E, Zavasnik-Bergant T, Begić L, Kos J. Cystatin C as a potential marker for relapse in patients with non-Hodgkin B-cell lymphoma. Cancer letters. 2007; 248: 192-7.
- Leto G, Sepporta MV. The potential of cystatin C as a predictive biomarker in breast cancer. Expert review of anticancer therapy. 2020; 20: 1049-56.
- Terpos E, Katodritou E, Tsiftsakis E, Kastritis E, Christoulas D, Pouli A, et al. Cystatin-C is an independent prognostic factor for survival in multiple myeloma and is reduced by bortezomib administration. Haematologica. 2009; 94: 372-9.
- Guo S, Xue Y, He Q, He X, Guo K, Dong P, et al. Preoperative serum cystatin-C as a potential biomarker for prognosis of renal cell carcinoma. PloS one. 2017; 12: e0178823.
- 35. Yang QK, Su YN, Wang W, Wang N, Yao ZX, Zhang XJ. CONUT Score or/and Peripheral Blood CD4+/CD8+ Ratio-Based Web Dynamic Nomograms to Predict the Individualized Survival of Patients with Advanced Osteosarcoma. Cancer management and research. 2020; 12: 4193-208.
- Huang X, Hu H, Zhang W, Shao Z. Prognostic value of prognostic nutritional index and systemic immune-inflammation index in patients with osteosarcoma. J Cell Physiol. 2019; 234: 18408-14.
- Sheen H, Kim W, Byun BH, Kong CB, Song WS, Cho WH, et al. Metastasis risk prediction model in osteosarcoma using metabolic imaging phenotypes: A multivariable radiomics model. PloS one. 2019; 14: e0225242.
- Tian MX, Liu WR, Wang H, Zhou YF, Jin L, Jiang XF, et al. Tissue-infiltrating lymphocytes signature predicts survival in patients with early/intermediate stage hepatocellular carcinoma. BMC medicine. 2019; 17: 106.
- Xu WH, Wu J, Wang J, Wan FN, Wang HK, Cao DL, et al. Screening and Identification of Potential Prognostic Biomarkers in Adrenocortical Carcinoma. Frontiers in genetics. 2019; 10: 821.
- Huang Y, Liu Z, He L, Chen X, Pan D, Ma Z, et al. Radiomics Signature: A Potential Biomarker for the Prediction of Disease-Free Survival in Early-Stage (I or II) Non-Small Cell Lung Cancer. Radiology. 2016; 281: 947-57.
- Zhao C, Li H, Wang L, Sun W. An Immunohistochemical Study of Stathmin 1 Expression in Osteosarcoma Shows an Association with Metastases and Poor Patient Prognosis. Medical science monitor: international medical journal of experimental and clinical research. 2018; 24: 6070-8.
- Jiang Y, Chen C, Xie J, Wang W, Zha X, Lv W, et al. Radiomics signature of computed tomography imaging for prediction of survival and chemotherapeutic benefits in gastric cancer. EBioMedicine. 2018; 36: 171-82.
- Granzier RWY, van Nijnatten TJA, Woodruff HC, Smidt ML, Lobbes MBI. Exploring breast cancer response prediction to neoadjuvant systemic therapy using MRI-based radiomics: A systematic review. European journal of radiology. 2019; 121: 108736.
- Fabbri M, Calin GA. Epigenetics and miRNAs in human cancer. Advances in genetics. 2010; 70: 87-99.

- Della Ragione F, Gagliardi M, D'Esposito M, Matarazzo MR. Non-coding RNAs in chromatin disease involving neurological defects. Frontiers in cellular neuroscience. 2014; 8: 54.
- Xin S, Wei G. Prognostic factors in osteosarcoma: A study level meta-analysis and systematic review of current practice. Journal of bone oncology. 2020; 21: 100281.